

Microevolution and Patterns of Transmission of *Shigella sonnei* within Cyclic Outbreaks of Shigellosis, Israel

Adi Behar, Kate Susan Baker, Ravit Bassal,
Analia Ezernitchi, Lea Valinsky,
Nicholas R. Thomson, Daniel Cohen

Whole-genome sequencing unveiled host and environment-related insights to *Shigella sonnei* transmission within cyclic epidemics during 2000–2012 in Israel. The Israeli reservoir contains isolates belonging to *S. sonnei* lineage III but of different origin, shows loss of tetracycline resistance genes, and little genetic variation within the O antigen: highly relevant for *Shigella* vaccine development.

Shigellosis is common all over the world and is hyperendemic to developing countries where children with the disease have an increased risk for persistent diarrhea, arrested growth, and death (1–3). The annual incident cases of shigellosis are estimated at ≈190 million in developing countries, where *Shigella flexneri* is the most common cause of shigellosis, and ≈1 million in industrialized countries, where *S. sonnei* predominates (4–7).

The Study

Despite the improved socioeconomic conditions, Israel has remained an area where shigellosis is highly endemic, reporting an annual incidence rate of culture-proven shigellosis of ≈97 cases per 100,000 population. Cyclic outbreaks during 2000–2012 occur every 2 years; *S. sonnei* is the pathogen for >85% of the cases. It has been shown that the ultraorthodox Jewish communities, which are overcrowded and have a high number of children <5 years of age, were the epicenter of these epidemics during the past 15 years (5). We used whole-genome sequencing (WGS) to provide a high-resolution view to better understand the local microevolution and patterns of *S. sonnei* transmission within the cyclic outbreaks in Israel.

Author affiliations: Tel Aviv University, Tel Aviv, Israel (A. Behar, D. Cohen); Kimron Veterinary Institute, Beit Dagan, Israel (A. Behar); The Wellcome Trust Sanger Institute, Hinxton, Cambridge, UK (K.S. Baker, N.R. Thomson); University of Liverpool, Liverpool, UK (K.S. Baker); Israel Center for Disease Control, Jerusalem, Israel (R. Bassal); Central Laboratories, Ministry of Health, Jerusalem (A. Ezernitchi, L. Valinsky)

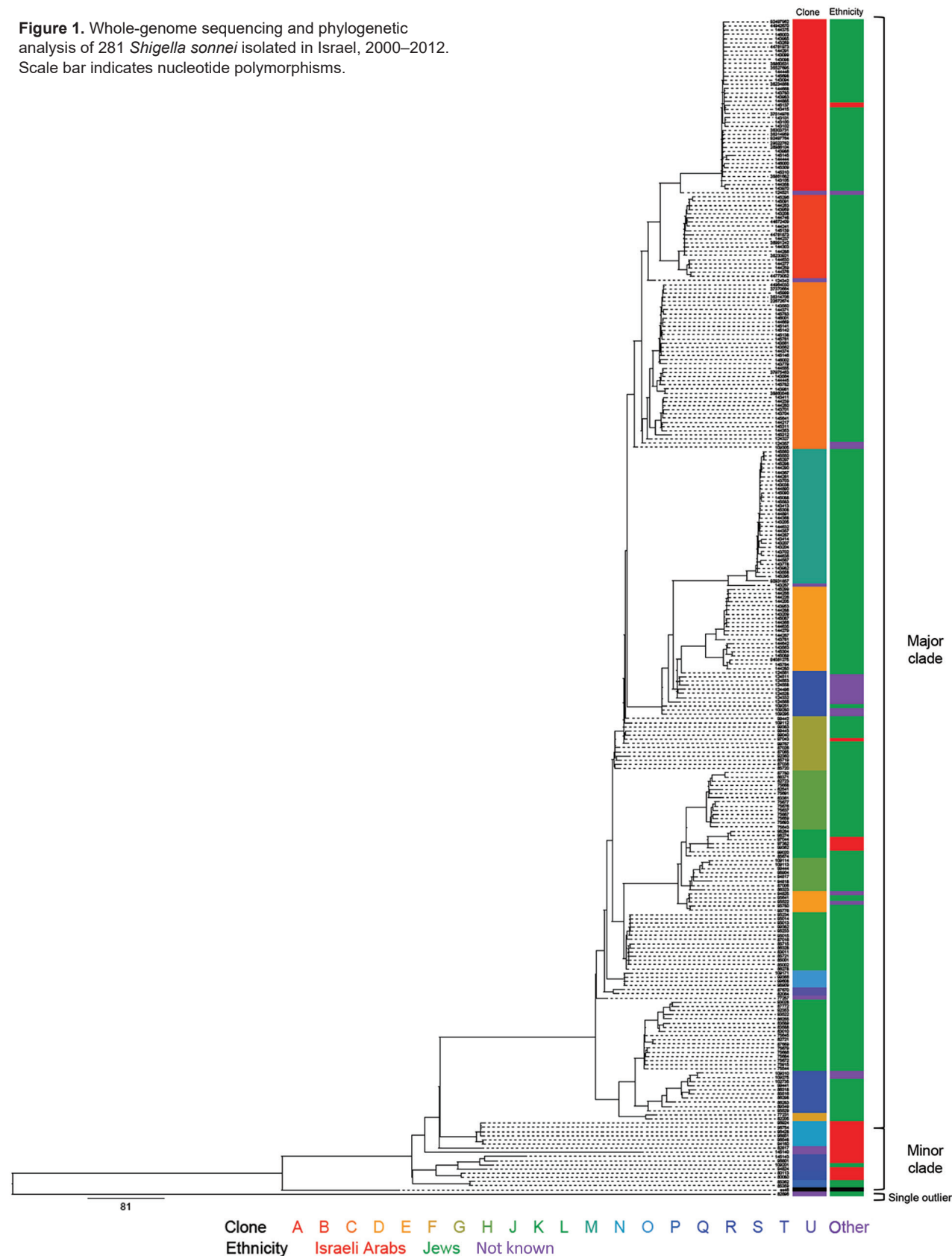
DOI: <https://doi.org/10.3201/eid2407.171313>

A total of 281 *S. sonnei* isolates were subject to WGS (Figure 1; online Technical Appendix Table 1, <https://www.cdc.gov/EID/article/24/7/17-1313-Techapp1.pdf>). We collected data from isolates during the epidemic years 2000, 2002, 2004, 2006, 2008, and 2012, and the nonepidemic years 2001 and 2003. All isolates were from children of various sanitary, socioeconomic, cultural, and ethnic backgrounds: ultraorthodox Jews, secular Jews, and Israeli Arabs. The ultraorthodox Jews represent ≈11% of the total population of Israel. This population group resides in towns or neighborhoods separated from the secular Jewish population (8) and also in mixed ones. The Israeli Arabs, who are estimated to account for 20% of the total population, reside mostly in rural areas and in towns or neighborhoods separated from the Jewish population; but they also live in towns inhabited by both Jews and Arabs (8). Of the 281 isolates, 263 (93.5%) were collected from Jewish children (mainly from ultraorthodox communities) and 18 (6.4%) isolates were from Israeli Arab children (mainly Bedouins living in southern Israel).

The WGS analysis showed that the clones within the Israeli reservoir formed 2 distinct subclades: a major subclade (subclade I) containing ≈94% of the Israeli collection, which is more prevalent among Jewish children (92% originated from Jewish children); and a minor subclade (subclade II) containing ≈5.7% of the Israeli collection, which is more prevalent among Israeli Arab children (82%). Only 1 isolate (≈0.3%) did not cluster with any of the Israeli isolates (Figure 1).

A comparison to global analyses (9) suggests that even though both subclades belong to *S. sonnei* lineage III, they are of different origins. Subclade II clones were more closely related to isolates that originated in Egypt and Iran than to the Israeli subclade I clones that seem to be endemic and have a distinctive recombination site, as previously described for 1 sequenced isolate from a patient in Israel in 2003 (9; online Technical Appendix Table 2). They were also found to distinguish *S. sonnei* among Jewish Orthodox communities of various countries (10). Nine of 13 Israeli Arab strains in clade II were isolated from Bedouins living in the vicinity of the Egyptian border. The frequent migration over the Israel–Egypt border of Bedouins often belonging to the same tribe

Figure 1. Whole-genome sequencing and phylogenetic analysis of 281 *Shigella sonnei* isolated in Israel, 2000–2012. Scale bar indicates nucleotide polymorphisms.



could explain the possible importation of subclade II *S. sonnei* from Egypt and/or through Egypt, similar to the recent transborder silent spread of poliovirus type 1, another fecal–orally transmitted enteropathogen in southern Israel (11). Our results also indicate that in general, isolates from Israeli Arab children who reside in mixed settlements and in close proximity to Jewish children commonly have positive test results for *Shigella* strains in clade 1. Only 5 (1.9%) isolates in clade I originated from Israeli Arabs (Figure 1). Of note, 4 of the 5 isolates were obtained from samples from Arab children residing

in Beer Sheva (3 isolates) and Mevaseret Zion (1 isolate), cities inhabited by both Jews and Arabs. Consequently, it appears that a combination of both biogeography and ethnicity forming microhabitats for *S. sonnei* clone circulation shapes the differences observed between Jewish and Israeli Arab children.

Each subclade could be further subdivided into clonal groups consisting of clusters of isolates with ≤ 30 chromosomal single-nucleotide polymorphism (SNP) differences from the nearest neighboring cluster. We defined a total of 20 unique and distinct *S. sonnei* endemic clones

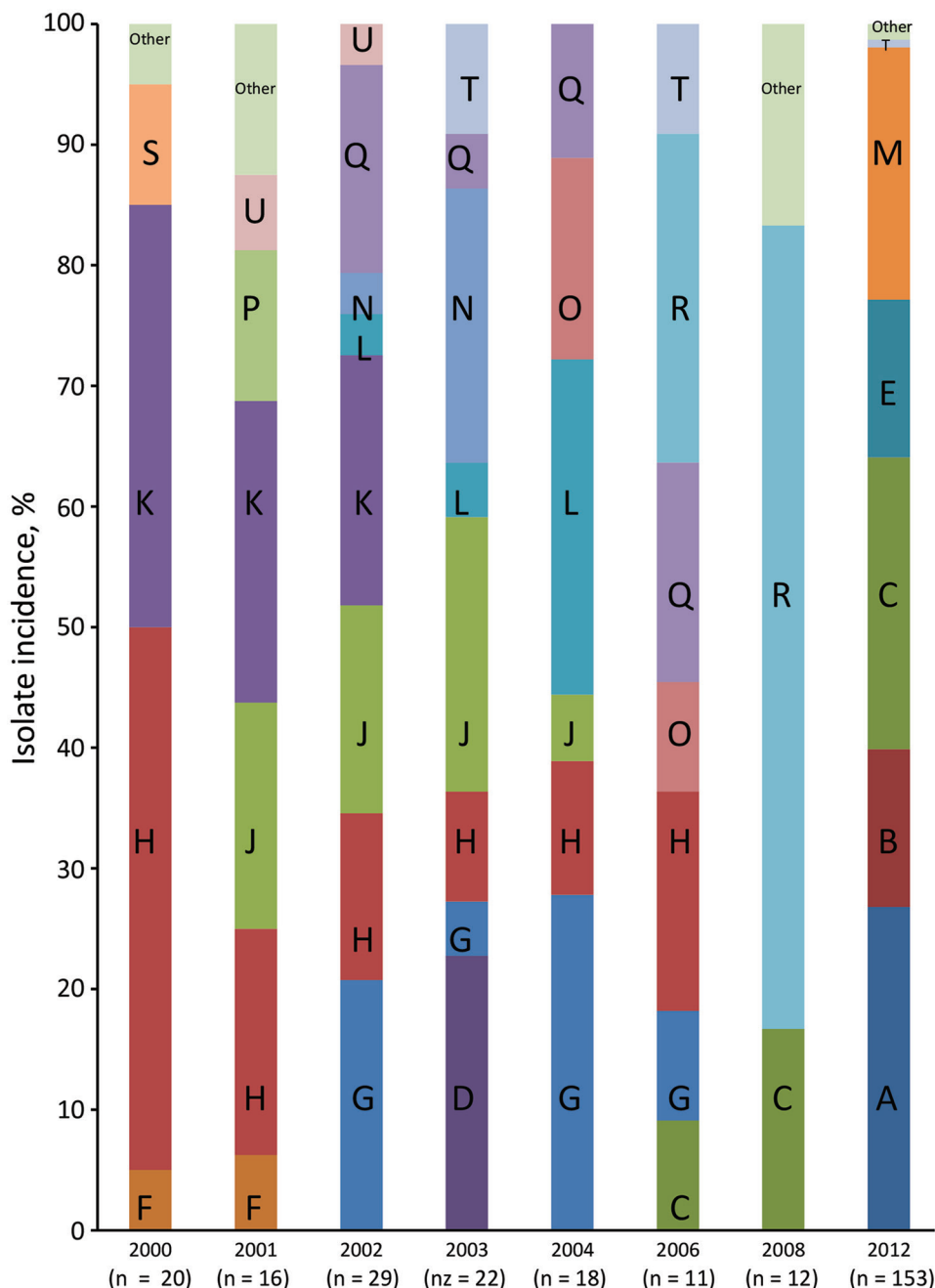


Figure 2. The relative distribution of the various whole-genome sequencing defined *Shigella sonnei* clones per year of isolation, 2000–2012.

circulating in the Israeli population (Figure 2, panels A–U). The majority of the clones ($\approx 69\%$; Figure 2) can be found throughout the years regardless of the shigellosis outbreaks that occurred in Israel every 2 years during 2000–2012, suggesting some mechanism of persistence (5). Contrary to our hypothesis, neither the establishment and dynamics of persistent or dominant clones could explain the Israeli cyclic outbreaks. Moreover, we found no specific genetic attributes that could distinguish them from other clones. Therefore, we postulate that the cyclic peaks of morbidity rates associated with *S. sonnei* are the result of changes in the level of natural immunity, as was shown by several observational studies (5,12,13). An outbreak of shigellosis occurring among children 0–4 years of age will lead to an increase in the level of natural immunity to the homologous *Shigella* organism (*S. sonnei*), which will also provide the level of herd immunity sufficient to prevent the onset of a new epidemic. After 1 or 2 years, declining levels of antibodies, together with the intake of a new cohort of naive newborns, will lead to a decrease in the level of herd immunity below a critical level. High and continuous exposure to a variety of circulating *S. sonnei* clones in children 0–4 years of age who live in crowded conditions will lead to the renewal of the epidemic transmission of these clones (5).

Conclusions

Although we excluded all *S. sonnei* plasmids from the phylogenetic analysis, plasmid reads were mapped and the assembled sequences compared with the reference plasmid sequences. Our data suggest that plasmid spA is undergoing degradation as a result of the loss of tetracycline resistance genes over time. This finding is consistent with the results of Holt et al. for the Middle East (III) clade (9) and with laboratory examination showing that the *S. sonnei* Israeli reservoir is becoming less resistant to tetracycline (5) (p-value for linear trend <0.01) (online Technical Appendix Table 3).

Although notoriously unstable when *S. sonnei* is grown on laboratory media, invasive plasmid pINVB was present in $\approx 58\%$ of our isolate sequences. Our results demonstrate that *S. sonnei* O antigen encoded on this plasmid is well-conserved within the *S. sonnei* Israeli reservoir. No SNPs were detected in genes that belong to the O antigen gene cluster in $\approx 97\%$ of the plasmids, and pINVB seems to be under very little immune selection as has been also shown in other studies (9,14). We identified a single SNP leading to a nonsynonymous substitution, in gene *wbgW* within the O antigen gene cluster that was shared by only 4 (2.4%) isolate plasmids. We also identified in 1 ($\approx 0.6\%$) isolate 1 SNP, a nonsynonymous change in gene *wbgY*. To date, *Shigella* vaccine development has mainly focused on serotype-

targeted vaccines that are based on *shigella* O antigen (15). Thus, our findings may have implications for public health as the need for a safe and effective *Shigella* vaccine becomes more pressing (15).

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About the Author

Dr. Behar is a researcher working at the Parasitology division at Kimron Veterinary Institute. During this research, she was a research fellow at the Department of Epidemiology and Preventive Medicine, School of Public Health, Sackler Faculty of Medicine, Tel Aviv University, Tel Aviv, Israel. Her research interests include the tripartite interactions between bloodsucking insects, their microbes, and the pathogens they transmit.

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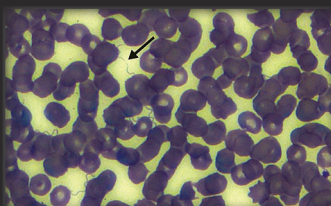
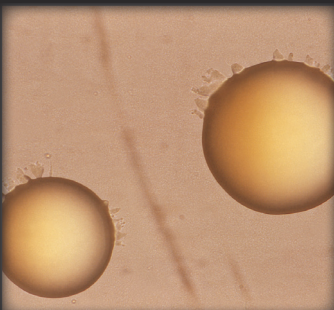
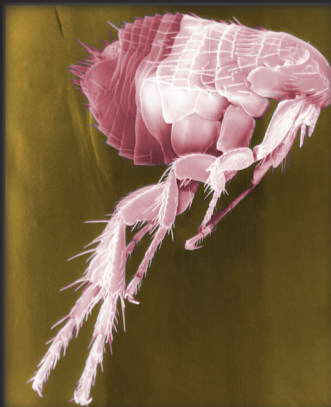
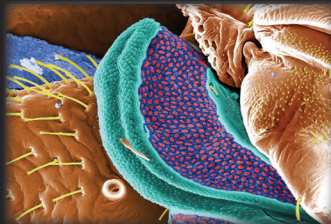
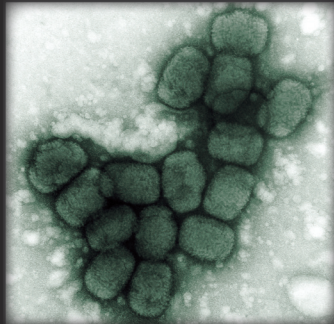
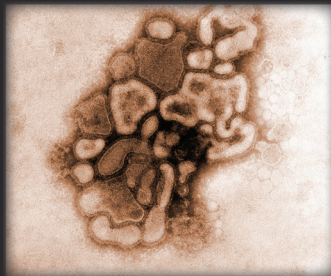
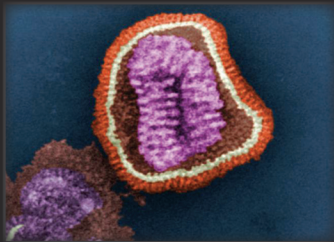
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Address for correspondence: Ali Behar, Kimron Veterinary Institute– Parasitology, PO Box 12, Beit Dagan 50250, Israel; email: adib@moag.gov.il

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Microevolution and Patterns of Transmission of *Shigella sonnei* within Cyclic Outbreaks of Shigellosis, Israel

Technical Appendix Part 1: The Methodology used in this Study for Whole-genome Sequencing and SNP-based Analysis

For whole genome sequencing, bacterial DNA from *S. sonnei* isolates was extracted from colonies by using Wizard Genomic DNA Purification Kit (Promega, Madison, WI, USA) according to the manufacturer's protocol. Index-tagged paired-end Illumina sequencing libraries were prepared, as previously described (1) and were sequenced on the Illumina Genome Analyzer GAII according to the manufacturer's protocols to generate tagged paired-end reads. Reads from each isolate were mapped to the *S. sonnei* reference genome (strain Ss046 chromosome: NC_007384; strain Ss046 plasmids: NC_007385, NC_009347, NC_009346 and NC_009345; plasmid pEG356: NC_013727) by using Bowtie (2) to produce a binary alignment map (BAM). SAMtools34 (3) was used to create a variant call format (VCF) file from each of the BAMs, which was further parsed to extract only single nucleotide polymorphism (SNP) positions which were of high quality in all genomes. Regions of unexpectedly high SNP density that might have been introduced by mobile element movement or recombination, were removed from the alignment by using gubbins (4), and a maximum-likelihood phylogeny was derived using RAxML (5). Gene content were examined manually using the comparative genomic approaches in Artemis and ACT (6). Wellcome Trust Sanger Institute sequence data are available in the Short Read Archive under the accession numbers provided in Technical Appendix Table 1.

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Technical Appendix Part 2: Israeli sub-clade positions within the global *S. sonnei* phylogeny determined by Holt et al. (7)

Technical Appendix Table 1. Accession numbers for the 281 isolates included in this study.

Identification number	Accession number
22672674	ERR190914
26986104	ERR190906
29022762	ERR190916
35527695	ERR190911
36230921	ERR190913
36234868	ERR190907
36303731	ERR190915
36314706	ERR190910
36314959	ERR190908
36860531	ERR190909
36860548	ERR190912
36861682	ERR190917
36991242	ERR190905
44773052	ERR211146
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37975463	ERR211148
94081275	ERR211149

Identification number	Accession number
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92497962	ERR211157
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75887	ERR190759
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80113	ERR190764
82064	ERR190767
82205	ERR190768
82541	ERR190769
82721	ERR190770
82723	ERR190771
82817	ERR190772
82896	ERR190773
83010	ERR190774
83011	ERR190775
83361	ERR190776
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85001	ERR190779
85002	ERR190780
85359	ERR190781
85362	ERR190782

Identification number	Accession number
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85719	ERR190786
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86328	ERR190795
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87016	ERR190797
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99362	ERR190841
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Identification number	Accession number
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145783	ERR319304
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146001	ERR319308
146002	ERR319309
146003	ERR319310

Technical Appendix Table 2. Similarity of the Israeli minor sub-clade isolates with global *S. sonnei* mapping by Holt et al. (7)

Name	Clone	Ethnicity	Isolation date	Similarity with isolates from:
80080	S	Israeli Arabs	2000	Egypt
80113	S	Israeli Arabs	2000	Egypt
85359	P	Jews	2001	Korea
85362	P	Jews	2001	Korea
82817	Other	Israeli Arabs	2001	UK
94160	N	Israeli Arabs	2002	Iran
96851	N	Israeli Arabs	2003	Iran
96924	N	Israeli Arabs	2003	Iran
96428	N	Israeli Arabs	2003	Iran
96548	N	Israeli Arabs	2003	Iran
96754	N	Israeli Arabs	2003	Iran
94824	T	Israeli Arabs	2003	Egypt
96801	T	Israeli Arabs	2003	Egypt
109201	T	Jews	2006	Egypt
145143	T	Israeli Arabs	2012	Egypt
145140	Other	Israeli Arabs	2012	UK
One isolate that did not clade with any of the Israeli strains (belong to Global II)				
82896	Other	Jews	2001	Brazil, Senegal

Technical Appendix Table 3. Resistance of *S. sonnei* isolates to tetracycline (2000–2012)

Year	Total number of isolates	Tetracycline-resistant isolates*	%
2000	20	20	100
2001	16	10	62.5
2002	29	8	27.5
2003	22	6	27
2004	18	3	17
2006	11	0	0
2008	12	0	0
2012	153	29	19

p for linear trend <0.01*